# Stereoselective Synthesis of *syn-* and *anti-*3a-Hydroxyl-1, 2, 3, 3a, 8, 8a-hexahydropyrrolo[2,3-*b*] indole-2-carboxylic Acid

Xiao Yin YANG<sup>1</sup>, Christian HAUG<sup>1, 2</sup>, Yi Ping YANG<sup>1</sup>, Zhi Sheng HE<sup>1</sup>, Yang YE<sup>1</sup>\*

<sup>1</sup>State Key Laboratory of Drug Research, Shanghai Institute of Materia Medica, Shanghai Institutes for Biological Sciences, Chinese Academy of Science, Shanghai 200031

<sup>2</sup>Current address: Bicoll Biotechnology (Shanghai) Co. Ltd., 518A Bibo Road, Shanghai 201203

**Abstract:** The structure of *syn-* and *anti-*N<sup>b</sup>-(*p*-toluenesulfonyl)-3a-hydroxyl-1, 2, 3, 3a, 8, 8a-hexahydro pyrrolo[2,3-*b*] indole-2-carboxylic *t*-butyl ester was stereoselectively synthesized by oxidative ring formation of N<sup>b</sup>-(*p*-toluenesulfonyl)-L-tryptophan *t*-butyl ester in methylene chloride containing dimethyldioxirane (DMDO), the *p*-toluenesulfonyl group and *t*-butyl group can be easily removed by sodium naphthalenide and trifluoroacetic acid, respectively.

Keywords: Oxidative ring formation, stereoselective synthesis, sodium naphthalenide.

Alkaloids with 3a-hydroxyl-1, 2, 3, 3a, 8, 8a-hexahydropyrrolo[2,3-b] indole-2carboxylic acid (HPI, **A**) substructure have been isolated from bacteria, fungi, plant and marine organism. These natural products show diverse biological activities, *e.g.* in antitumor, acyl-CoA (cholesterol acyltransferase) inhibition, neuropeptide and cholecystokinin receptor antagonistic inhibition. HPI is regarded as their toxophoric unit<sup>1</sup>. Thus the synthesis of this key substructure has been attracting the interest of organic chemists, some procedures towards the synthesis of HPI have been developed<sup>2</sup>. However these procedures are relatively complicated and a by-product, N-formylkynurenine **B**<sup>3</sup>, was usually produced in these procedures. Recently the first diastereoselective approach to the *syn-* and *anti*-HPI scaffolds using the different configurations of D- and L-tryptophane as the starting material was reported<sup>4</sup>. Herein, we report a newly elaborated steroselective route for the synthesis of *syn-* and *anti*-HPI by using L-tryptophane as sole starting material.



<sup>\*</sup>E-mail: yye@mail.shcnc.ac.cn

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Our research started from the naturally abundant L-tryptophane 1. *p*-Toluenesulfonyl-L-tryptophane 2 was synthesized smoothly from L-tryptophane 1 referring to the former literature<sup>5</sup>. Compound 3 was obtained by treating p-toluenesulfonyl-L-tryptophane 2 with N, N'-diisopropyl-O-tert-butylisourea without racemization under inert gas. Treatment of 3 with N-bromosuccinimide (NBS) and triethylamine generated the unstable dihydropyrroindole 5, which could be quickly purified by chromatography on silica gel. Subsequent treatment of this compound with 3,3-dimethyl-dioxirane (DMDO) at -78°C followed by reduction with sodium borohydride led to the key intermediate 6 substantially as a single diastereomer (3aS, 8aR). According to the <sup>1</sup>H-NMR spectra data the ratio of diastereomers is 41.2 : 2.7 (d.e. = 88%). This diastereometric excess can be enhanced by a purification over silica gel to d.e. > 95%. The relative configuration was determined by X-ray analysis and <sup>1</sup>H-NMR, <sup>13</sup>C-NMR. The syn-compound **4** can also be easily obtained by direct oxidation of 3 with DMDO at -78°C in 92% yield and d.e. of 72%. The configuration of 4 was determined by comparing of nmr data with compound 6( see Scheme 1).

It is well known that the *p*-toluenesulfonyl protecting group attached to unaromatic amino group is quite difficult to be removed. In our approach, sodium naphthalenide was

Scheme 1



Reagents and conditions: (a) Ts-Cl, NaHCO<sub>3</sub> 95%; (b) N, N'-diisopropyl-O-*tert*- butylisourea, dry  $CH_2Cl_2$ , 85%; (c) NBS,  $Et_3N$ , 91%; (d) DMDO, -78°C; (e) ( i ) DMDO, -78°C; (ii ) NaBH<sub>4</sub>, 97%; (f) NaC<sub>10</sub>H<sub>8</sub>, DME, 65%; (g) TFA,  $CH_2Cl_2$ , 98%.

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employed to cleave this group to yield compound **7** in 65% with dimethoxyethane (DME) as solvent <sup>6</sup>. As to the *t*-butyl protecting group, it is quite easy to remove quantitatively in the presence of trifluoroacetic acid to afford the target molecular **8** (see **Scheme 1**).

The structures of compound **2**, **3**, **4**, **6**, **7** and *anti*-HPI **8** were confirmed by <sup>1</sup>H-NMR, <sup>13</sup>C-NMR, ESIMS<sup>7</sup>.

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- Selected data of compound 6: Colorless needle, <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 600 MHz, δ<sub>ppm</sub>): 1.10(s, 9H, *t*-Bu), 2.40(s, 3H, Ts-CH<sub>3</sub>), 2.47(dd, 1H, J=9.7Hz, J=13.0Hz, H-3), 2.69(dd,1H, J=1.3Hz, J=13.0 Hz, H-3), 4.33(dd, 1H, J=1.3 Hz, J=9.7 Hz, H-2), 5.00 (s, 1H,H-8a), 6.62(d, 1H, J=7.8Hz, H-7), 6.80(t, 1H, J=7.8Hz, H-5), 7.18(m, 2H, H-4, 6), 7.36(d, 2H, J=7.3Hz, Ts), 7.80 (d, 2H, J=7.3Hz, Ts); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100 MHz, δ<sub>ppm</sub>): 21.5(q,Ts-CH<sub>3</sub>), 27.3(q×3, *t*-Bu), 40.6(t, C-3), 61.6 (d, C-2), 81.8(s, C-3a), 83.5 (d, C-8a), 87.6(s, *t*-Bu), 110.8(d, C-7), 119.8(d, C-5), 123.8(d, C-3b), 127.1(d×2, Ts), 128.5(s, Ts), 129.9(d×2, Ts), 131.1(d, C-6), 135.7(s, C-3b), 144.0(C-7a), 149.8(d, Ts), 169.4(d, COO-*t*-Bu). ESIMS (*m*/*z*): 431.0(25%) [M+H]<sup>+</sup>, 453.1(100%) [M+Na]<sup>+</sup>.

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